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Patentanmeldung Nr. | Patent application No. | Demande de brevet n°

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Der Präsident des Europäischen Patentamts;
Im Auftrag

For the President of the European Patent Office

Le President de l'Office européen des brevets
p.o.

R.C. van Dijk

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Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
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An efficient system for RNA silensing

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An efficient system for RNA silencing

Field of the invention

The invention relates to a method for efficient RNA silencing in eucaryotic cells, particularly plant cells. Consequently, the method can be used to reduce the phenotypic expression of an endogenous gene in a plant cell. Furthermore the method can be applied in a high throughput screening for RNA silencing.

Background of the invention

RNA silencing is a type of gene regulation based on sequence-specific targeting and degradation of RNA. The term encompasses related pathways found in a broad range of eukaryotic organisms, including fungi, plants, and animals. In plants, RNA silencing serves as an antiviral defense, and many plant viruses encode suppressors of silencing. Also it becomes clear that elements of the RNA silencing system are essential for gene regulation in development. The emerging view is that RNA silencing is part of a sophisticated network of interconnected pathways for cellular defense, transposon surveillance, and regulation of development. Based on the sequence specific RNA degradation, RNA silencing has become a powerful tool to manipulate gene expression experimentally. RNA silencing was first discovered in transgenic plants, where it was termed co-suppression or posttranscriptional gene silencing (PTGS). Sequence-specific RNA degradation processes related to PTGS have also been found in ciliates, fungi, and a variety of animals from *Caenorhabditis elegans* to mice (RNA interference). A key feature uniting the RNA silencing pathways in different organisms is the importance of double-stranded RNA (dsRNA) as a trigger or an intermediate. The dsRNA is cleaved into small interfering RNAs (21 to 25 nucleotides) of both polarities, and these are thought to act as guides to direct the RNA degradation machinery to the target RNAs. An intriguing aspect of RNA silencing in plants is that it can be triggered locally and then spread via a mobile silencing signal. In plants, RNA silencing is correlated with methylation of homologous transgene DNA in the nucleus. Other types of epigenetic modifications may be associated with silencing in other organisms.

It is known from the art that transgenes encoding ds or self-complementary (hairpin) RNAs of endogenous gene sequences are highly effective at directing the cell's degradation mechanism against endogenous (ss) mRNAs, thus giving targeted gene

suppression. This discovery has enabled the transgenic enhancement of a plant's defense mechanism against viruses that it is unable to combat unaided. It has also shed light on how antisense and co-suppression might operate: by the inadvertent integration of two copies of the transgenes in an inverted repeat orientation, such that read-through transcription from one gene into the adjacent copy produces RNA with self-complementary sequences.

RNA silencing is induced in plants by transgenes designed to produce either sense or antisense transcripts. Furthermore, transgenes engineered to produce self-complementary transcripts (dsRNAs) are potent and consistent inducers of RNA silencing. Finally, replication of plant viruses, many of which produce dsRNA replication intermediates, causes a type of RNA silencing called Virus Induced Gene Silencing (VIGS). Whether VIGS, and the different types of transgene-induced RNA silencing in plants result from similar or distinct mechanisms is still a matter of debate. However, recent genetic evidence raises the possibility that the RNA silencing pathway is branched and that the branches converge in the production of dsRNA.

Until recently RNA silencing was viewed primarily as a thorn in the side of plant molecular geneticists, limiting expression of transgenes and interfering with a number of applications that require consistent, high-level transgene expression. With our present understanding of the process, however, it is clear that RNA silencing could have enormous potential for engineering control of gene expression, as well as for the use as a tool in functional genomics. It could be experimentally induced and targeted to a single specific gene or even to a family of related genes. Likewise, ds RNA-induced TGS may have similar potential to control gene expression. Although several methods for RNA silencing have been described in the art (WO99/53050, WO99/32619, WO99/61632, and W098/53083), there is clearly a need to develop alternative and more efficient tools for RNA silencing. In the present invention we have developed a highly efficient method for RNA silencing that can also be used as a tool for high throughput silencing. Said method uses a host that carries already a silenced locus and a second recombinant gene comprising a region that is homologous with the silenced locus. Although it is known from the art that the recombinant gene will be silenced, we have surprisingly found that also target genes, which have no significant homology with the silenced locus but have homology with the recombinant gene, are efficiently silenced.

Figure legends

Fig. 1: Schematical outline of homology between a silenced locus X, a recombinant gene Y and a target gene Z.

Fig. 2: Schematical outline of the T-DNA constructs that are present in silenced locus X_1, recombinant gene Y_1 and target gene Z_1 (T-DNAs of pGVCHS287, pGUSchsS and pXD610 respectively) and of the transcript homology between X_1, Y_1 and Z_1.

LB and RB: left and right T-DNA border respectively; Pnos: nopaline synthase promoter; hpt: hygromycin phosphotransferase coding sequence; 3'nos: 3'untranslated region of the nopaline synthase gene; P35S; Cauliflower mosaic virus 35S promoter; nptII c.s., neomycin phosphotransferase II coding sequence; 3'chs: 3'untranslated region of the chalcone synthase gene of Anthirrinum majus; +1: transcription start; A_n: poly A-tail; gus c.s.: β-glucuronidase coding sequence; Pss: promoter of the small subunit of rubisco; bar: phosphinotricine transferase coding sequence; 3'g7: 3'untranslated region of the Agrobacterium octopine T-DNA gene 7; 3'ocs: 3'untranslated region of octopine synthase gene.

Fig. 3: Schematical outline of the T-DNA construct present in silenced locus X_1 and of the transiently introduced T-DNAs Y_2 (T-DNAs of pGVCHS287 and pPs35SCAT1S3chs, respectively) and of the transcript homology between X_1, Y_2 and Z_2 (the catalase1 endogene). Abbreviations as in Fig. 2

Fig. 4: Schematical outline of the T-DNA constructs present in silenced locus X_2 and of the transiently introduced T-DNAs Y_2 (T-DNAs of pGUSchsS + pGUSchsAS, and pPs35SCAT1S3chs, respectively) and of the transcript homology between X_2, Y_2 and Z_2 (the catalase1 endogene). Abbreviations as in Fig. 2

Detailed description of the invention

The present invention deals with an efficient method for RNA silencing in an eucaryotic host. The method makes use of a host that already comprises a silenced locus. Such a silenced locus can be generated by methods known in the art. For example the publication of De Buck and Depicker, 2001 and other publications, and also patents WO99/53050, WO99/32619, WO99/61632, and W098/53083 describe methods to

obtain RNA silencing. The 'target gene' is here defined as the gene of interest for silencing or to down-regulate its expression. An important aspect of this invention is that said target gene has no significant homology with the silenced locus. No significant homology means that either the overall homology is less than 40, 35, 30, 25% or even less, or that no contiguous stretch of at least 23 identical nucleotides are present (Thomas et al., 2001). Homology is typically measured using sequence analysis software (e.g., Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, Wis. 53705). Such software matches similar sequences by assigning degrees of homology to various insertions, deletions, substitutions, and other modifications. Silencing of said target gene in the present invention occurs via an intermediate step and hence our method is designated as domino silencing (Fig. 1). In said intermediate step a recombinant gene construct is introduced by transformation into the host comprising the silenced locus. Said recombinant gene construct has a region of homology with the silenced locus already present. Said region of homology is preferably more than 60, 70, 80, 90, 95 or even more than 99% homologous. The homologous region between the silenced locus and said recombinant gene can be found in the 5' untranslated or 3' untranslated region of the recombinant gene construct. Furthermore, said recombinant gene construct has a region of minimal 23 nucleotides (Thomas et al., 2001), but preferably longer, that are identical with the target gene, or has a region of overall homology of more than 60, 70, 80, 90, 95 or even more than 99%.

Thus in one embodiment the invention provides a method for obtaining efficient RNA silencing of a target gene comprising the introduction of a recombinant gene into a host that comprises a silenced locus and an unsilenced target gene whereby said recombinant gene comprises a region that is homologous with said silenced locus and whereby said target gene has homology with said recombinant gene but has no significant homology with said silenced locus.

In another embodiment the method is used wherein said host is a plant.

In another embodiment the method of the invention can be used for high throughput gene silencing. Indeed, a gene library can be made wherein for example every gene or coding region thereof is combined with a region of homology with the silenced gene that resides in the silenced locus.

In yet another embodiment the invention provides a plant that comprises a silenced locus and wherein a silenced target gene is obtained through the introduction of a recombinant gene according to the current method of the invention.

In yet another embodiment the RNA silencing of the target gene is obtained in more than 80, 85, 90 or 95% of the transgenic organisms.

In yet another embodiment the RNA silencing of the target gene occurs at an efficiency of more than 80, 85, 90 or 95 % as compared to the level of the unsilenced expression of the target gene.

Examples

A posttranscriptionally silenced inverted repeat transgene locus can trigger silencing of a reporter gene producing non-homologous transcripts.

We studied the interaction between three transgene loci X_1, Y_1 and Z_1 (Fig. 2, For a detailed description of all loci and constructs, see materials and methods) to address the question whether or not a stepwise homology between loci can lead to silencing.

It has been demonstrated previously that the posttransciptionally silenced *nptII* genes in locus X_1 are capable to in trans silence transiently expressed genes with partial transcript homology to their *nptII* transcripts (Van Houdt et al., 2000 b). We subsequently found that also a stably expressed β-glucuronidase (*gus*) gene (in locus Y_1), with partial transcript homology to the *nptII* transcripts of the silencing inducing locus X_1, becomes efficiently silenced in trans (Fig. 2: X_1 and Y_1 and table 1: X_1Y_1 compared to Y_1). On the contrary, the *nptII* genes of locus X_1 are not able to trigger silencing of the gus genes in locus Z_1 which is expected as the genes of both loci produce transcripts without significant homology (Fig. 2). The homology between the two transcripts of X_1 and Y_1 is mainly situated in the 3'untranslated region (250 nucleotides), but also the 5'untranslated sequences show a small region of homology (29 nucleotides). These results demonstrate that the in trans silencing effects are not triggered by promoter homology. When Y_1 and Z_1 loci are combined in so called Y_1Z_1 hybrids both types of gus genes, having transcript homology in the gus coding sequence of 1809 nucleotides, remain highly expressed as reflected in the normal gus activity showing that the RNA silencing mechanism does not become activated (Table 1: Y_1Z_1 compared to Y_1 and Z_1). Interestingly however, upon creation of a stepwise homology between X_1 and Z_1 by introducing locus Y_1, the new observation described here is that also the gus expression in locus Z_1 is reduced in $X_1Y_1Z_1$ plants (Table 1:

$X_1Y_1Z_1$ compared to Y_1Z_1). Thus, creating a stepwise homology between a silenced locus and a target gene by introducing a recombinant gene is sufficient to trigger silencing of the target.

Silencing inducing transgene loci can trigger silencing of a non-homologous endogene. We further assessed the universality and the usefulness in high throughput functional gene analyses of silencing elicited by a stepwise homology in trans, called domino silencing. Therefore, we evaluated whether the expression of the tobacco endogenous catalase1 (cat1) genes is reduced in plants carrying a silencing locus (X locus) showing no significant homology with the catalase endogene by introducing a recombinant gene (Y construct). As silencing locus we used either X_1 or X_2 (Fig.2: locus X_1, Fig.3: locus X_2), in either case containing the 3' chalcone synthase sequences of Anthirrinum majus (3'chs). As transmitter for silencing we constructed a recombinant gene composed of the catalase1 coding sequence and the 3' chs region under control of the 35S promoter (P35S) (residing on T-DNA pPs35SCAT1S3chs, Fig.2 and 3: T-DNA in Y_2). The recombinant cat1 3'chs genes (Y2) were introduced in tobacco leaves bearing locus X_1 (or X_2) via Agrobacterium injection. As a negative control, we introduced a recombinant gene in which the cat1 coding sequence is replaced by the gus coding sequence (pGUSchsS, T-DNA construct as in locus Y_1 Fig.1). In this case, no stepwise homology is created between the silencing inducing locus and the target catalase endogenes. As a positive control, the recombinant construct Y_2 was also introduced in transgenic tobacco with silenced catalase1 genes by the presence of a catalase1 antisense construct (Cat1AS in Champnongpol et al., 1996). Sixteen days after Agrobacterium injection, the catalase activity was determined in protein extracts of injected leaf tissue and compared with the activity in non-injected wild type (SR1) leaf tissue (Table 2). The results indicate that domino silencing is also applicable to endogenes since the catalase activity is clearly reduced in 6 out of 7 samples, while it remains high in the negative controls. In conclusion, not only an inverted repeat-bearing silencing-inducing transgene locus, but also a silencing-inducing locus in which the two residing chimeric genes give rise to transcripts with complementarity in the 3'UTR (3'chs)(Fig.3: X_2), is able to trigger domino silencing reducing endogenous catalase expression.

Table 1: Results of a GUS-activity determination in protein extracts of leaf tissue harvested from tobacco plants containing different combinations of the loci X_1, Y_1 and Z_1 (Fig.2). The mean values of a number of plants (n) are given.

genotype	GUS-act.[1] 4 weeks[2] U GUS/mg TSP	n	GUS-act. Mature[2] U GUS/mg TSP	n
X_1	<[3]	1	<	1
Y_1	368 ± 165[4]	9	n.d.	-
Z_1	126 ± 30	10	48 ± 8	5
X_1Y_1	2 ± 1	4	4 ± 2	4
X_1Z_1	139 ± 35	9	46 ± 14	5
Y_1Z_1	477 ± 101	10	231 ± 106	6
$X_1Y_1Z_1$[5]→ Y_1Z_1	195 ± 104	16	315 ± 46	8
→ $X_1Y_1Z_1$	4 ± 3	22	12 ± 4	9

[1] The mean GUS-activity (GUS-act.) was calculated, using n samples and expressed as units (U) GUS per miligram of total soluble protein (TSP).

[2] The plants were analyzed in two different developmental stages; 4 weeks after sowing and at a mature stage just before onset of flowering.

[3] below detection limit (1 U GUS/mg TSP)

[4] standard deviation

[5] Growth of $X_1Y_1Z_1$ plants was performed in conditions that both Y_1Z_1 and $X_1Y_1Z_1$ plants were able to develop. A PCR screen with X_1-specific primers was performed to discriminate between presence and absence of X_1.

n.d. not determined

Table 2: Results of a catalase-activity determination in protein extracts of leaf tissue harvested from Agrobacterium injected tobacco leaves.

Genotype injected plant	Construct introduced via Agrobacterium injection	catalase activity 16 days after injection (60 µg TSP)
WT (SR1)	- (non-injected)	-0.2116[2] 100%[3]

X_1	pGUSchsS	-0.2556 121%
X_1	Y_2	-0.0589 27%
X_1[4]	Y_2	-0.0698 33%
X_2	pGUSchsS	-0.1782 84%
X_2	Y_2	-0.0641 30%
X_2	Y_2	-0.0987 47%
X_2[4]	Y_2	-0.0914 43%
X_2[4]	Y_2	-0.1996 94%
X_2[4]	Y_2	-0.0627 30%
Cat1AS	Y_2	-0.0439 21%

[1] X_1, see Fig. 3; X_2, see Fig. 4

[2] the mean of two samples independently measured (-0.2270 and –0.1963)

[3] The catalase activity in wild type SR1 tobacco leaves was set to 100 %.

[4] 24 hours after Agrobacterium injection, the plants were placed under high light conditions for 24 hours (1000 µmol / m^2 s). This treatment is known to stimulate endogenous catalase 1 transcription. As the degree of cat suppression is similar in uninduced as in induced situation, the data indicate that enhanced transcription of the endogenous catalase target is not required to trigger domino silencing.

Materials and Methods

Plasmid construction

pPs35SCAT1S3chs: The T-DNA of this plasmid is schematically shown in Fig. 2 :Y'.

Description of the transgene loci and production of hybrid plants

Locus X_1 harbours an inverted repeat about the right T-DNA border of construct pGVCHS287, carrying a neomycinphosphotransferase II (*nptII*) gene under the control of the Cauliflower mosaic virus 35S promoter (P35S) and the 3'signalling sequences of the Anthirrinum majus chalcone synthase gene (3'chs). The *nptII* genes are posttranscriptionally silenced and can trigger in trans silencing and methylation of homologous target genes (Van Houdt et al., 2000 a and b and Fig.2).

Locus Y_1 contains a single copy of the pGUSchsS T-DNA, containing a gus gene under the control of P35S and 3'chs (in transformant GUSchsS29) and shows normal levels of gus expression (Fig.2).

Locus Z_1 contains more than one copy of the pXD610 T-DNA, harbouring the gus gene under control of P35S and the 3'untranslated region (UTR) of the nopaline synthase gene (3'nos), (in plant LXD610-2) and shows normal gus expression (De Loose et al., 1995 and Fig.2).

Locus X_2 contains a single copy of both the pGUSchsS and pGUSchsAS T-DNA (in transformant GUSchsS+GUSchsAS 11) and triggers silencing in cis of the gus genes, but also in trans of (partially) homologous genes (Fig.4).

X_1 and Z_1 hemizygous plants were obtained as hybrid progeny of the crossing of tobacco plants homozygous for locus X_1 (=HOlo1; Van Houdt et al., 2000 a and b) and homozygous for locus Z_1 (=LXD610-2/9 De Loose et al., 1995) to wild type SR1 respectively. Y_1 hemizygous plants were obtained by crossing the hemizygous primary tobacco transformant GUSchsS29 to SR1 and selecting for the presence of locus Y_1 in the hybrid progeny. X_1Y_1 and Y_1Z_1 hemizygous plants are the hybrid progeny plants of the cross between Holo1 and GUSchsS29 and between GUSchsS29 and LXD610-2/9 respectively that are selected for the presence of Y_1. X_1Z_1 hemizygous plants are the hybrid progeny of the cross between Holo1 and LXD610-2/9. $X_1Y_1Z_1$ hemizygous plants were obtained by crossing X_1Y_1 hemizygous plants to LXD610-2/9; as we only selected for the presence of Y_1 in the hybrid progeny both Y_1Z_1 and $X_1Y_1Z_1$ hemizygous plants were obtained.

Preparation of Agrobacteria and injection

The Agrobacteria C58C1Rif^R(pGV2260)(pGUSchsS)Cb^R,PPT^R or C58C1Rif^R(pMP90) (pPs35SCAT1S3chs)Gm^R,PPT^R were mainly grown as described by Kapila et al., 1997 exept that the Agrobacteria were resuspended in MMA to a final OD_{600} of 1. Greenhouse grown plants of 10 to 15 cm in height were used. Half of the third top leaf was injected via the lower surface using a 5ml syringe while the leaf remained attached to the plant. The plants were kept in the greenhouse and 16 days after injection three to four discs of 11 mm in diameter were exised from the injected tissue for the preparation of a fresh protein extract to determine the catalase activity.

Enzymatic assays

Preparation of the protein extracts and GUS-activity measurements were done as previously described (Van Houdt et al., 2000 b). Preparation of the protein extracts for catalase-activity measurement and the spectophotometric catalase-activity determination was done according to Champnongpol et al., 1996.

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Claims

1. A method for obtaining efficient RNA silencing of a target gene comprising the introduction of a recombinant gene into a host that comprises a silenced locus and a target gene whereby said recombinant gene comprises a region that is homologous with said silenced locus and whereby said target gene has homology with said recombinant gene but has no significant homology with said silenced locus.

2. A method according to claim 1 wherein said host is a plant.

3. A method according to claims 1 or 2 to obtain high through-put gene silencing.

4. A plant comprising a silenced target gene obtainable by a method according to claims 1 or 2.

5. A method according to claims 1 or 2 wherein said RNA silencing of the target gene is obtained in more than 95% of the hosts.

6. A method according to claims 1 or 2 wherein RNA silencing of the target gene is obtained in more than 85% of the hosts.

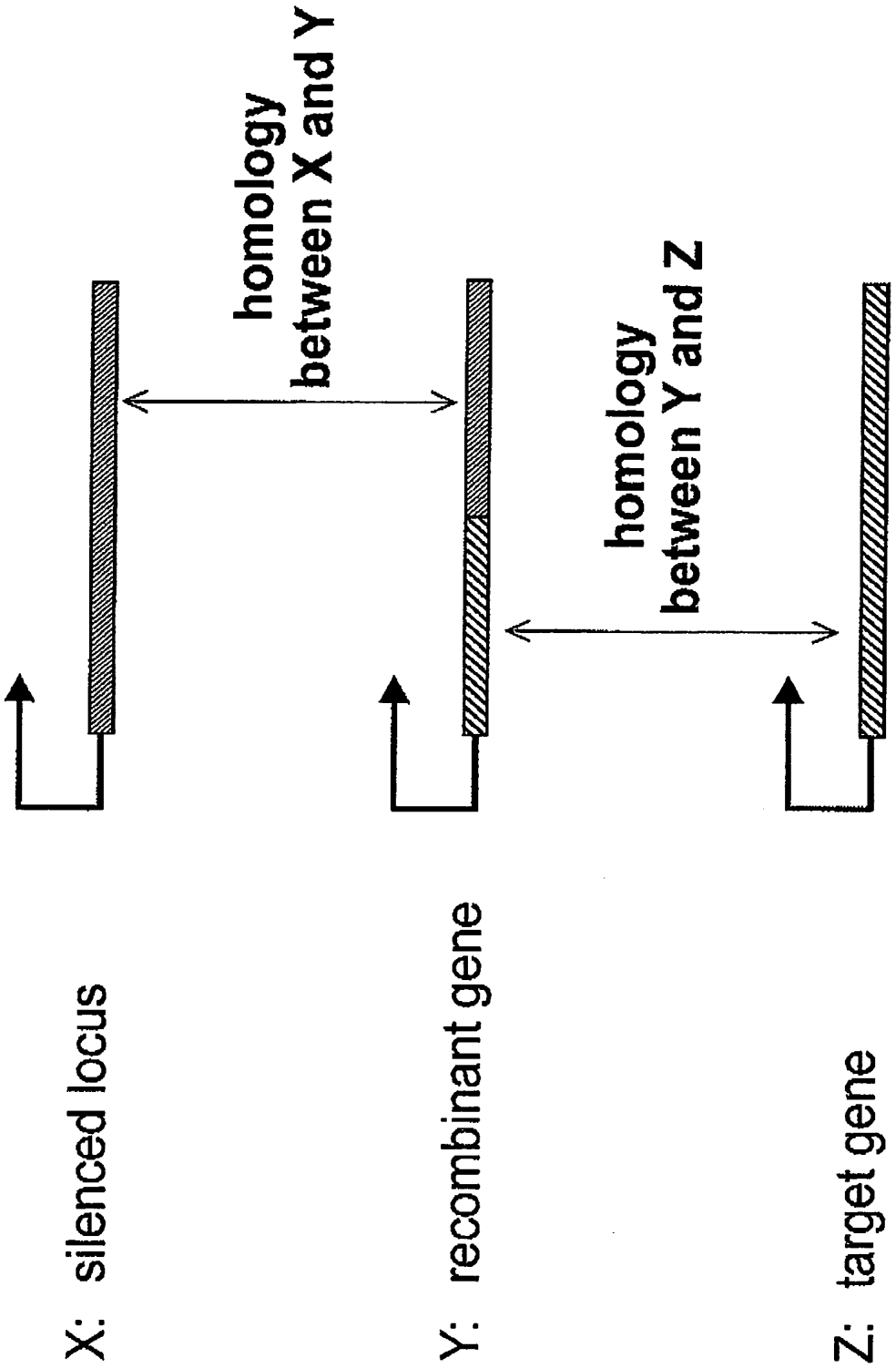
7. A method according to claims 1 or 2 wherein said RNA silencing of the target gene occurs at an efficiency of more than 95 % as compared to the level of the unsilenced expression of the target gene.

8. A method according to claims 1 or 2 wherein said RNA silencing of the target gene occurs at an efficiency of more than 85 % as compared to the level of the unsilenced expression of the target gene.

Abstract of the invention

The invention relates to a method for efficient RNA silencing of target genes in eucaryotic cells, particularly plant cells. Consequently, the method can be used to reduce the phenotypic expression of an endogenous gene in a plant cell. Furthermore the method can be applied in a high throughput screening for mutant phenotypes as a result of RNA silencing of any endogene.

Figure 1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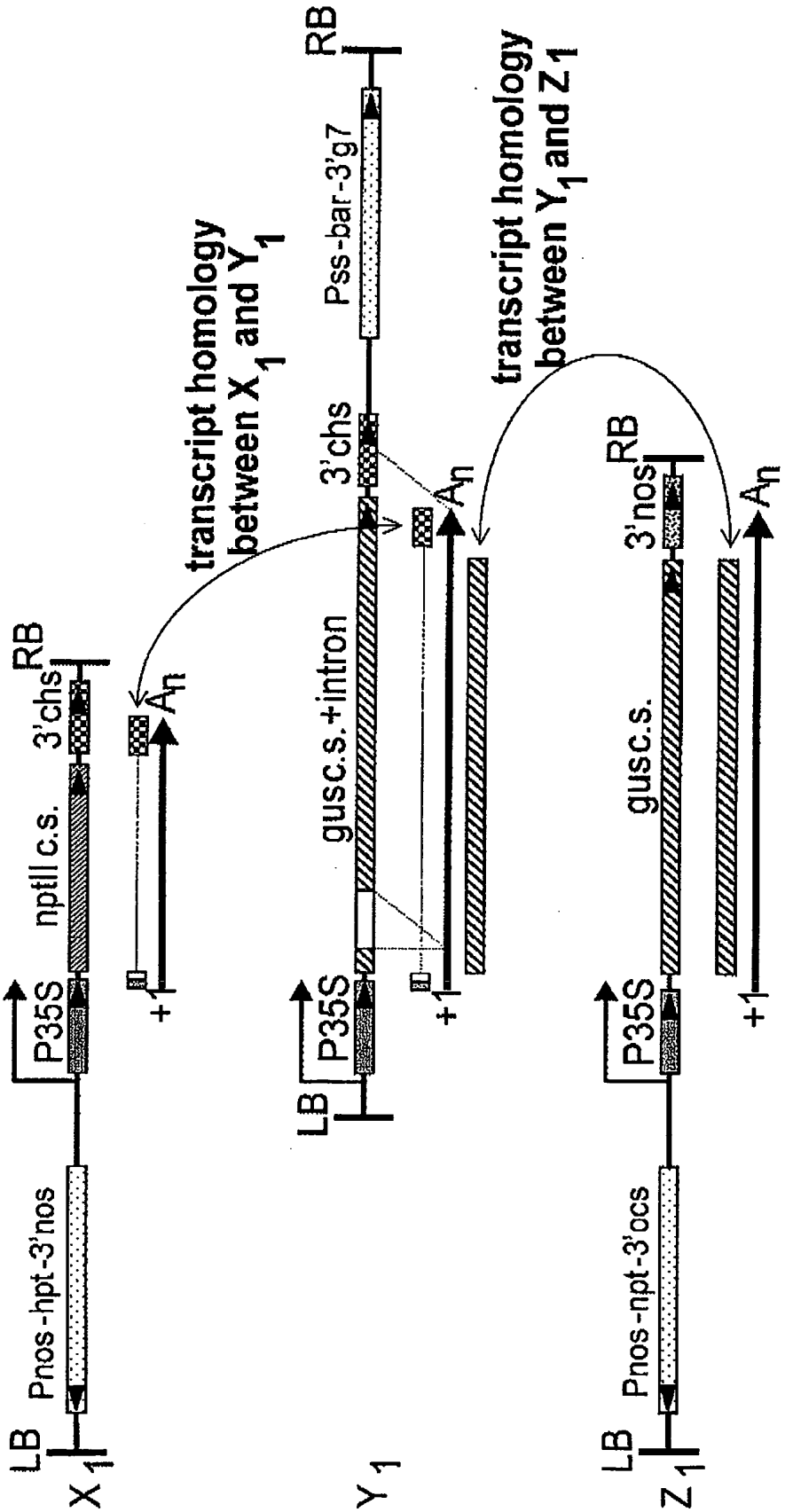
between X and Y
homology
between Y and Z
X: silenced locus
Y: recombinant gene
Z: target gene

Figure 2


transcript homology between X1 and Y1
transcript homology between Y1 and Z1
LB
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P35S
nptII c.s.
3'chs
RB
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+1
X1
LB
P35S
gusc.s.+intron
3'chs
Pss-bar-3'g7
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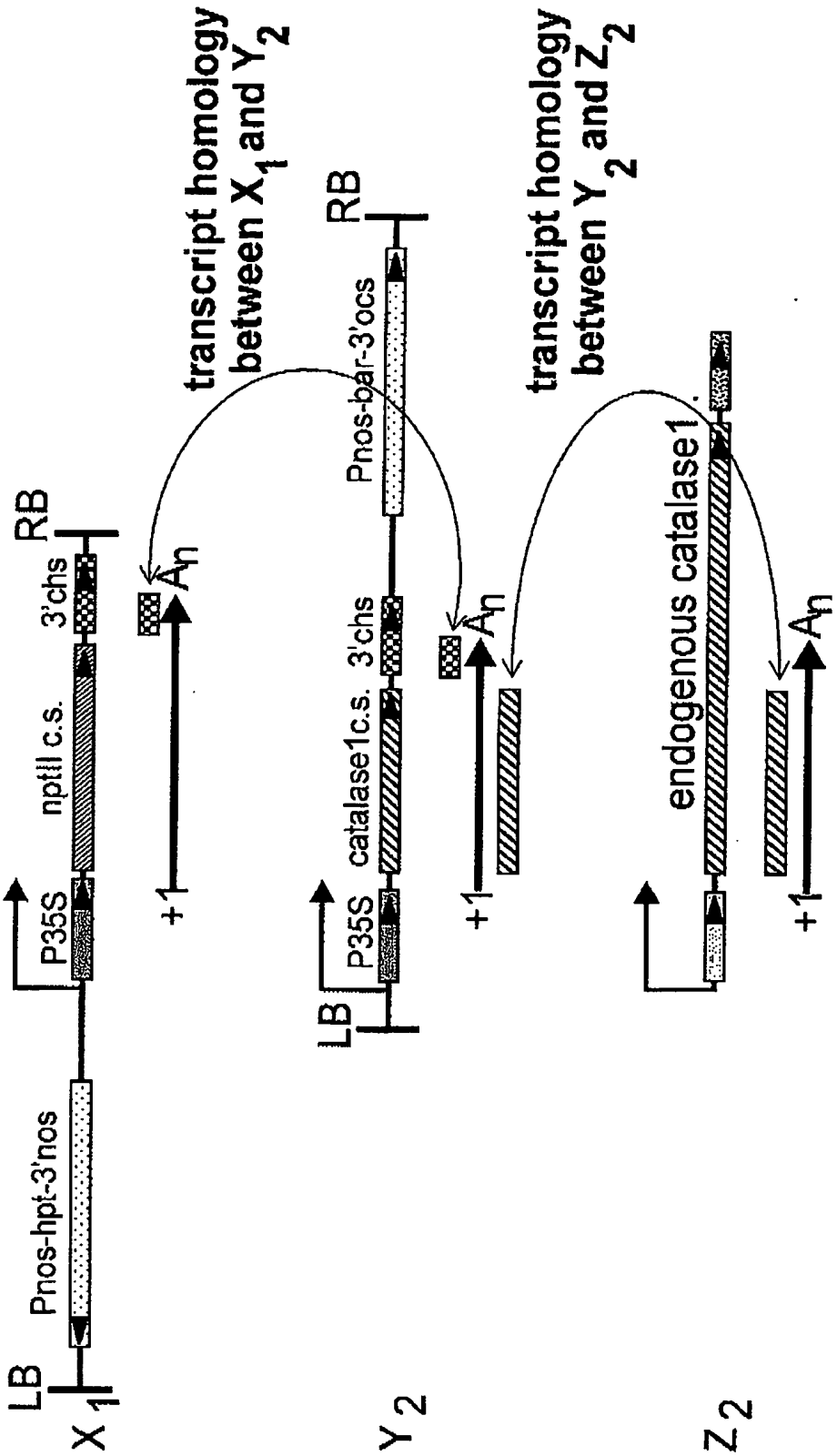
+1
Z1

Figure 3


transcript homology between X_1 and Y_2
transcript homology between Y_2 and Z_2
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Pnos-hpt-3'nos
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nptII c.s.
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X_1
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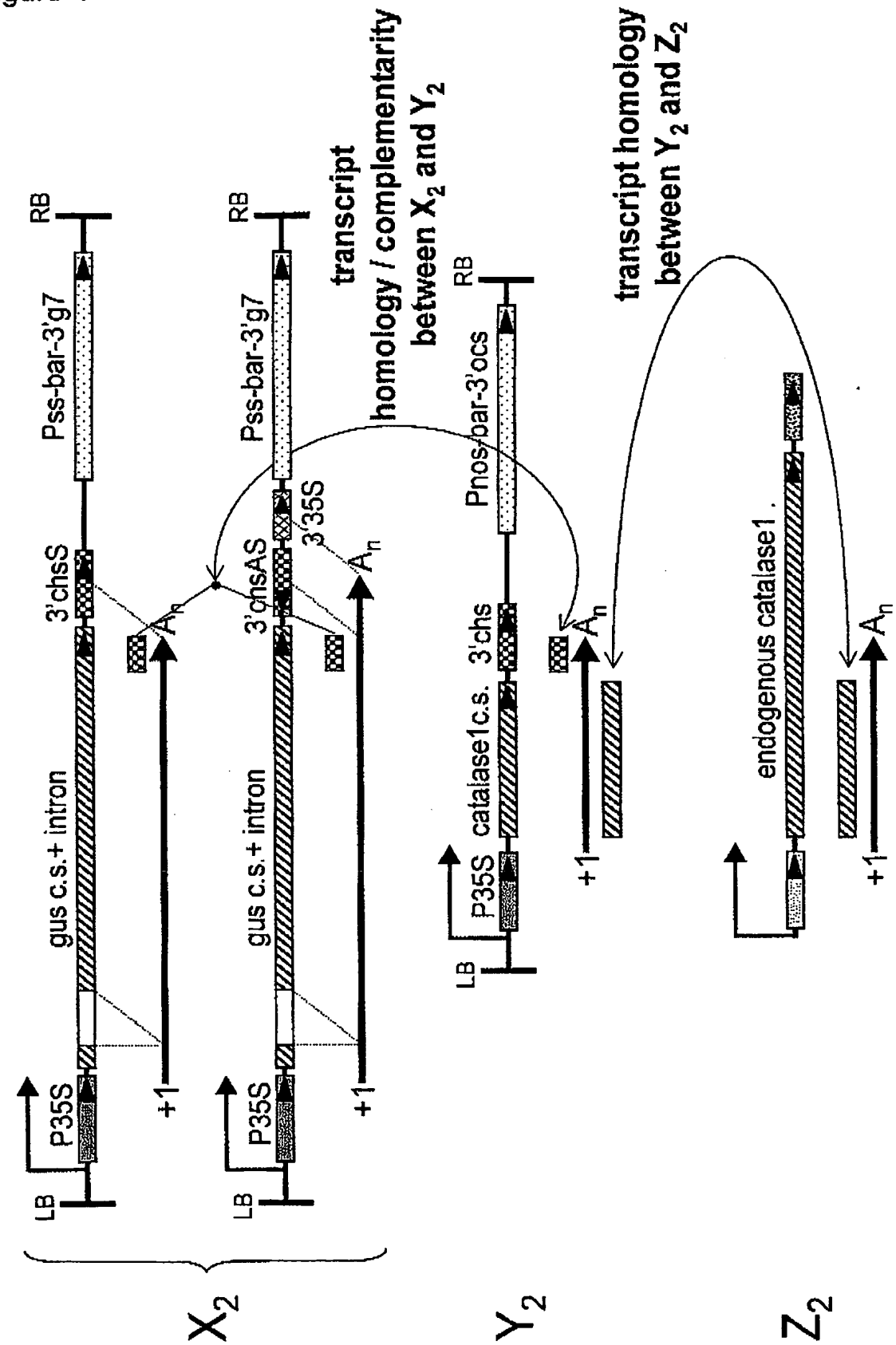
Y_2
endogenous catalase1
Z_2

Figure 4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Pnos-bar-3'ocs
3'chs
catalase1c.s.
transcript homology between Y_2 and Z_2
endogenous catalase1 .
X_2
Y_2
Z_2